

Review: Management of Postprandial Diarrhea Syndrome

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ABSTRACT

Unexpected, urgent, sometimes painful bowel movements after eating are common complaints among adults. Without a clear etiology, if pain is present and resolves with the movements, this is usually labeled “irritable bowel syndrome-diarrhea” based solely on symptoms. If this symptom-based approach is applied exclusively, it may lead physicians not to consider treatable conditions: celiac disease, or maldigestion due to bile acid malabsorption, pancreatic exocrine insufficiency, or an α -glucosidase (sucrase, glucoamylase, maltase, or isomaltase) deficiency. These conditions can be misdiagnosed as irritable bowel syndrome-diarrhea (or functional diarrhea, if pain is not present). Limited testing is currently available to confirm these conditions (antibody screens for celiac disease; fecal fat as a surrogate marker for pancreatic function). Therefore, empirical treatment with alpha amylase, pancreatic enzymes, or a bile acid-binding agent may simultaneously treat these patients and serve as a surrogate diagnostic test. This review will summarize the current evidence for bile acid malabsorption, and deficiencies of pancreatic enzymes or α -glucosidases as potential causes for postprandial diarrhea, and provide an algorithm for treatment options.

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Currently, the diagnosis of irritable bowel syndrome is based predominantly upon symptoms listed in **Table 1**, recurrent abdominal pain coinciding with changes in stool frequency or consistency. The symptom-based diagnosis stems from validation studies that contrasted symptoms with “organic” diseases. For example, in the classical paper of Manning et al,¹ there was a comparison of the symptoms of 32 irritable bowel syndrome patients and 33 with organic diseases: 13 duodenal ulcer; 5 inflammatory bowel disease; 4 gastroesophageal reflux; 2 gastric ulcer, gallstones, and carcinoma of the colon respectively; and 5 miscellaneous gastrointestinal disorders. Primary care physicians have been advised to make a positive, symptom-based diagnosis of irritable bowel syndrome,² although the diagnostic accuracy of the symptom-based criteria has not been validated in

large patient cohorts where all structural organic diseases, or more subtle digestive disorders, had been definitively excluded.³

Although 50% of patients classified as having irritable bowel syndrome complain of postprandial diarrhea, symptom criteria for irritable bowel syndrome do not include the meal association.⁴ In addition, there are physiological changes stimulated postprandially, such as intestinal secretion, and ileocolonic and proximal colonic transit that may contribute to symptoms including bloating, diarrhea, or urgency.⁵⁻⁷

The diagnostic work-up for these patients typically focuses on identifying medical conditions with easily detectable abnormalities such as Crohn disease, celiac disease, occult neoplasm, or microscopic colitis. Once these are excluded, physicians may diagnose irritable bowel syndrome based on symptoms. Without further testing of such recommendations, patients also may receive centrally acting medications (such as antidepressants) in an attempt to control the central perception of abnormal gastrointestinal functions. Conditions that may produce symptoms that mimic irritable bowel syndrome, such as bile acid malabsorption,

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maldigestion due to pancreatic insufficiency, food allergy, or starch/sugar maldigestion due to glucosidase insufficiency or inhibition, are not usually considered.^{8,9} Consequently, patients may continue suffering, as illustrated in the case discussed below.

CASE REPORT

The patient, a 43-year-old woman, experienced episodes of diarrhea for approximately 11 years. They occurred 2-3 times weekly, approximately 2-4 hours after eating meals containing spices, corn, green beans, lima beans, oranges, cheese, salads, milk products, tomatoes, or garlic. Symptoms included cramping, which resolved after 3-5 watery, loose stools. Eating late might cause nocturnal attacks. Multiple family members, including her mother, had similar digestion problems. The patient was referred to a gastroenterologist. Colonoscopy, colon biopsies for microscopic colitis, serology for celiac disease, and small bowel follow-through were all normal. Therefore, the gastroenterologist diagnosed “irritable bowel syndrome-diarrhea” and recommended a probiotic. Because symptoms continued, her internist (author MEM) did additional tests: a Sudan stool test for fat (positive) and a fecal elastase-1 test screening for pancreatic exocrine insufficiency (normal). Subsequently, pancrelipase (PES) was prescribed before eating any “trigger meal.” This treatment eliminated the symptoms. The patient has had no digestion problems since using PES “on demand” for the last 2 years.

DISCUSSION

The differential diagnosis of postprandial diarrhea symptoms includes a rapid colonic response to feeding,⁴ food

allergies or hypersensitivity, food intolerance, altered gut flora or small bowel bacterial overgrowth, fructose maldigestion,¹⁰⁻¹³ and malabsorption. Food elimination diets or prolonged fasting may reduce symptoms to some degree.¹⁴⁻¹⁷ Such patients may undergo tests to exclude malabsorption and to identify the underlying cause. When standard testing for celiac disease, Crohn disease, and microscopic colitis is negative, physicians may assume the patient is suffering a form of functional diarrhea or irritable bowel syndrome-diarrhea, and the therapeutic options offered are limited, often to the current mainstream treatments used in irritable bowel syndrome-diarrhea (Table 2).

However, there are 3 specific classes of disease causing postprandial diarrhea that respond to targeted therapy; in these patients, the therapeutic trial may serve as a surrogate diagnostic test when sensitive and specific tests are not available, as might occur in the primary care setting.

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Pancreatic Exocrine Insufficiency

Recently, pancreatic exocrine insufficiency was detected in 6.1% of patients who fulfilled the Rome II criteria for irritable bowel syndrome-diarrhea by measuring the stool concentration of fecal elastase-1 (FEC).¹⁸ Frequently, diarrhea in diabetic patients is considered as being due to an autonomic defect, but pancreatic insufficiency has been identified by the presence of a reduced FEC concentration in both Type 1 and Type 2 diabetes in 56.7% and 35%, respectively.¹⁹ In another study, 21.1% of 546 patients with diabetes mellitus requiring insulin were found to have pancreatic exocrine insufficiency based upon a FEC <100 μg/g.²⁰ The sensitivity of the FEC test (55%) compared with the secretin-erulein test (which requires duodenal intubation and aspiration of duodenal contents following hormonal stimulation) for determining pancreatic exocrine sufficiency has been questioned. Measurement of FEC may not accurately diagnose mild pancreatic exocrine insufficiency, and a therapeutic trial of PES may still be helpful.²¹

Glucosidase Inhibition and Deficiency

Carbohydrates comprise a large percentage of most diets. Carbohydrates in the diet are complex entities and must be broken down into monosaccharides before they can be absorbed. Digestion starts with oral salivary amylase, followed by gastric amylase in the stomach until the amylase (which has a pH optimum of 7²²) becomes inactive due to the low intragastric pH as a result of

CLINICAL SIGNIFICANCE

- A careful history of what precedes diarrhea is important.
- Postprandial diarrhea suggests a maldigestion problem: bile acid malabsorption, pancreatic insufficiency, or a glucosidase deficiency or inhibition.
- If diagnostic studies (including celiac antibodies) are negative, empirical treatment with alpha amylase, pancreatic enzymes, or bile acid binders (taken immediately before eating) is appropriate.
- Effectiveness of the above empiric treatment suggests a maldigestion condition.

Table 1 Rome III Criteria for Irritable Bowel Syndrome⁸

The Rome III criteria, based only on symptoms, require that a patient experience recurrent abdominal pain or discomfort for at least 3 days per month in the last 3 months associated with 2 or more of the following:

Improvement with a bowel movement(s).

Onset of symptoms was associated with a change in the frequency of the stool(s) or

Onset of symptoms was associated with a change in the appearance of the stool(s).

From: Camilleri M. Do the symptom-based, Rome criteria of irritable bowel syndrome lead to better diagnosis and treatment outcomes? The con argument. *Clin Gastroenterol Hepatol*. 2009;8(2):129.⁸

Table 2 Current Treatment Recommendations for Irritable Bowel Syndrome-Diarrhea²

Bulking fiber agents
Lactase supplementation
Probiotics, antibiotics
Smooth muscle relaxants
Opioids: loperamide, diphenoxylate-atropine, deodorized tincture of opium
Serotonergic agents: 5-HT ₃ antagonist (eg, alosetron)
Antidepressants: tricyclic agents
Bile acid binders: cholestyramine, colestevlam

From: Lacy BE, Weiser K, De Lee R. The treatment of irritable bowel syndrome. *Therap Adv Gastroenterol*. 2009;2(4):221-238.²

stomach acid secretion, and continues with small intestinal brush border glucosidases.

The glucosidases are divided into α -glucosidases: maltase, isomaltase, sucrase, glucoamylase, and trehalase; and a β -glucosidase: lactase. Lactase deficiency is a well-recognized cause of diarrhea, although it has been demonstrated that the load of lactose is a significant factor in determining whether an individual who is lactase-deficient will actually develop diarrhea. Thus, when lactose intake is limited to the equivalent of 240 mL or less of milk a day, symptoms are likely to be negligible in individuals who identify themselves as severely lactose-intolerant.²³ Symptoms identical to those of lactose intolerance may occur when starch is incompletely digested due to an inhibition or deficiency of α -glucosidases.

There is considerable research being conducted on the relationship between the phytochemicals (ie, chemical compounds that occur naturally in plants, and may affect health, but are not established as essential nutrients) in food and their effect on the human body. These substances may potentially reduce the incidence of cancer, cardiovascular disease, diabetes, and osteoporosis.²⁴ Over 1200 different plant species have been recognized as possible hypoglycemic agents by inhibiting amylase or glucosidases.²⁵ Food extracts with this effect have included berries, vegetables (including beans, corn, eggplant, and pumpkin), and some colored grains.²⁶ In vitro studies have demonstrated glucosidase inhibition with cinnamon extract,²⁷ certain tropical pepper spices,²⁸ basil extract,²⁹ certain strains of maize,³⁰ certain Indian spices,³¹ the Welsh onion,³² the Maitake mushroom (*Grifola frondosa*),³³ and chamomile tea.³⁴ A consequence of glucosidase inhibition includes abdominal symptoms: bloating and postprandial diarrhea. Drugs marketed to reduce hyperglycemia and complications of diabetes: acarbose (Precose; Bayer Healthcare Pharmaceuticals Inc., West Haven, Conn), miglitol (Glyset; Pfizer, New York, NY), and voglibose are associated with gastrointestinal side effects. In the STOP-Noninsulin Dependent Diabetes Mellitus international trial from 1995 to 2001, 29.5% of patients assigned to acarbose, compared with 18.2% using placebo, discontinued the drug because of adverse gastrointestinal tract effects: flatulence, diarrhea, and ab-

dominal pain.³⁵ Quercetin, which is found in high concentrations in onions and garlic, is 5 times more potent than acarbose in α -glucosidase inhibition.³⁶

Because the following food triggers identified by patients: spicy foods, onions, garlic, corn, and peppers, inhibit glucosidases in vitro, it is conceivable that the resultant postprandial diarrhea is due to inhibition of either amylase or an α -glucosidase.

Congenital Glucosidase Deficiencies. Glucosidase deficiency is recognized predominantly in children with congenital sucrase-isomaltase deficiency, a rare autosomal recessive genetic condition with which children cannot digest starch.³⁷ The prevalence of congenital sucrase isomaltase deficiency is estimated to be only 1 in 5000 patients of European descent, compared with 1 in 20 among those from Alaska, Canada, and Greenland. However, 2%-9% of European Americans have a partial deficiency in the enzyme activity due to heterozygosity for the gene mutation.³⁸ Other glucosidase deficiencies have been identified. A glucosylase deficiency was identified in 3% of children with chronic diarrhea.³⁹ In 2002, Karnsakul et al reported that approximately 21% of children with recurrent abdominal pain syndrome (the equivalent of irritable bowel syndrome in adults), have an α -glucosidase deficiency that is not congenital sucrase isomaltase deficiency, and these children are unable to fully digest normal starches.⁴⁰

Studies have identified adults with varied digestive problems, including intermittent postprandial diarrhea, to be deficient in one or more glucosidases.⁴¹⁻⁴³ Small bowel biopsies in adult patients with diarrhea have significantly lower levels of lactase, maltase, and sucrase when compared with those of patients with functional dyspepsia.⁴⁴ Steatorrhea has been reported with glucosidase deficiency,⁴⁵ consistent with the classical studies showing that steatorrhea may occur when diarrhea is induced by an osmotic laxative in healthy volunteers.⁴⁶ A recent study showed significant improvement in symptoms and quality of life in patients with diarrhea-predominant irritable bowel syndrome when prescribed a very low carbohydrate diet for 4 weeks.⁴⁷ It is unclear whether those patients had a subclinical form of glucosidase malabsorption.

Bile Acid Malabsorption

Bile acid malabsorption is significantly under-recognized and may account for approximately 30% of diarrhea previously diagnosed as irritable bowel syndrome-diarrhea or chronic functional diarrhea.⁴⁸ Unfortunately, confirmatory testing requires stool collection and the measurement of whole body retention of ⁷⁵Se-homocholeic acid taurine, a gamma-emitting conjugated bile salt homologue, which is not available in the US. Although an intermediate in bile acid biosynthesis, 7 α -OH-4-cholesten-3-one leaks from the hepatocyte into plasma in direct proportion to the rate of bile acid synthesis; measurement of this key intermediate is available in only a few academic centers at the current time.

Even when the $^{75}\text{SeHCAT}$ testing is available, only a small fraction of patients get tested.⁴⁹ It has recently been recognized that bile acid malabsorption may result from an overproduction of bile acids due to a defect in the feedback regulation of hepatocyte bile acid synthesis from the ileal hormone, fibroblast growth factor 19.⁵⁰ This reduced feedback regulation results in increased bile acid synthesis in the hepatocytes, increased passage of bile acids into the colon, and diarrhea because of the secretory effects of dihydroxy bile acids.

Even if there is no absolute increase in bile acids, their presence may cause diarrhea by affecting starch digestion. New research has identified that bile salts can bind to starch, which may inhibit starch digestion by pancreatic enzymes.⁵¹

Alternative Treatment Options for Postprandial Diarrhea

Because the 3 conditions discussed above are difficult to prove, empirical treatment with an over-the-counter alpha amylase supplement, PES, or a bile acid sequestrant is a reasonable option, especially when specific testing is not available, and when exclusion of “trigger” dietary factors cannot be identified or their exclusion does not solve the clinical problem.

How Effective is PES for Postprandial Diarrhea? PES is effective in reducing the diarrhea of patients with irritable bowel syndrome among patients with proven reduced fecal elastase as a marker of pancreatic insufficiency.¹⁸ In addition, PES improved digestion in normal subjects consuming a fatty meal.⁵² PES also reduced bile acid malabsorption in patients with cystic fibrosis^{53,54} and chronic pancreatitis,⁵⁵ and improved fat malabsorption due to cystic fibrosis,⁵⁶ human immunodeficiency virus,⁵⁷ and the mild steatorrhea associated with the irritable bowel syndrome.⁵⁸ A large double-blind study comparing PES to placebo among patients with otherwise unexplained postprandial symptoms who have normal pancreatic exocrine function has not been conducted to date.

In a small double-blind, placebo-controlled, crossover pilot study involving 49 patients with irritable bowel syndrome-diarrhea, there was a reduction in postprandial symptoms among the irritable bowel syndrome-diarrhea patients who were willing to consume their “trigger meals.” The primary end point was the number of patients who chose PES as the effective drug. In an intention-to-treat analysis, 30/49 (61%) patients chose PES ($P = .078$) as the effective drug. Analysis of patients who identified PES as the effective drug demonstrated a reduction ($P \leq .001$) of the following symptoms: bloating, cramping, borborygmi, urgency to defecate, and number and consistency of stools compared with the period when they received placebo.⁵⁹

In a large internal medicine practice, PES has been used as first-line therapy for patients with postprandial diarrhea for the last 10 years. In a recent retrospective review of all patients with irritable bowel syndrome treated by one physician (author MEM), 104/144 patients received PES: me-

dian age 60 years (range: 17-95 years); 70% were female. Duration of symptoms ranged from 1 to 50 years; frequency of postprandial diarrhea ranged from daily to once monthly. Eighteen (17%) either never tried the medication or were lost to follow-up. Of the 86 patients with follow-up information, 71 (82.5%) reported that PES improved or eliminated their symptoms, while 15 (17.4%) reported no improvement. Approximately 15% reported mild constipation that was managed by a reduction in the frequency or amount of PES; 1 individual had an exacerbation of asthma; and 3 had worsening abdominal pain. After a median 3.7 years, 43 (50%) patients still take PES as needed upon “demand.” Special testing was conducted with equal frequency in the responders and nonresponders: colonoscopies in 55 of 71 (77.5%) positive responders and 11 of 15 (73%) nonresponders; testing for celiac disease in 39 of 71 (54.9%) responders and 9 of 15 (60%) nonresponders. All of these tests were normal. Some patients are now using an over-the-counter enzyme supplement containing α -amylase with equal success at a considerable cost savings.

Why are Pancreatic Enzymes Effective? The mechanism for this apparent efficacy of PES in postprandial diarrhea is unknown unless there is pancreatic insufficiency as proven by a low fecal elastase-1 value. However, amylase, a component of PES, has been shown to augment glucosidases,⁶⁰ and any fat malabsorption that results from rapid transit or osmotic diarrhea (up to 14 grams of fat can be expected to be malabsorbed per day^{46,61}) is potentially ameliorated by the lipase in PES. PES has not been shown to substantially affect gastrointestinal motility unless pancreatic insufficiency is present.^{62,63} Direct measurements of pancreatic enzyme secretion in response to a meal have not been made.

How Effective is Bile Acid Sequestrant Therapy? An extensive review of 18 studies on the incidence of bile acid malabsorption in patients with irritable bowel syndrome-diarrhea based upon $^{75}\text{SeHCAT}$ retention testing was published in 2009. Therapy with a bile acid binder resulted in 96% improvement in the 10% of patients with severe bile acid malabsorption ($\text{SeHCAT} < 5\%$), 80% improvement in the 32% with moderate ($^{75}\text{SeHCAT} < 10\%$), and 70% improvement in the 26% with mild malabsorption ($^{75}\text{SeHCAT} < 15\%$).⁶⁴ Habba⁶⁵ reported a 68% improvement in diarrhea with bile acid sequestrant therapy among patients labeled as having irritable bowel syndrome-diarrhea. The bile acid sequestrant, colesevelam, reduces the accelerated ascending colon transit in patients with irritable bowel syndrome-diarrhea;⁶⁶ the transit rate in the ascending colon is a determinant of stool consistency.

Some foods do not cause the diarrhea syndrome and may actually ameliorate these symptoms. Sun-dried raisins have been shown to lower total fecal bile acid concentration by a reduction in lithocholic and deoxycholic acid.⁶⁷ In addition, an extract of wine from the Jacquez grapes has been shown to inhibit the castor oil-induced diarrhea in mice.⁶⁸

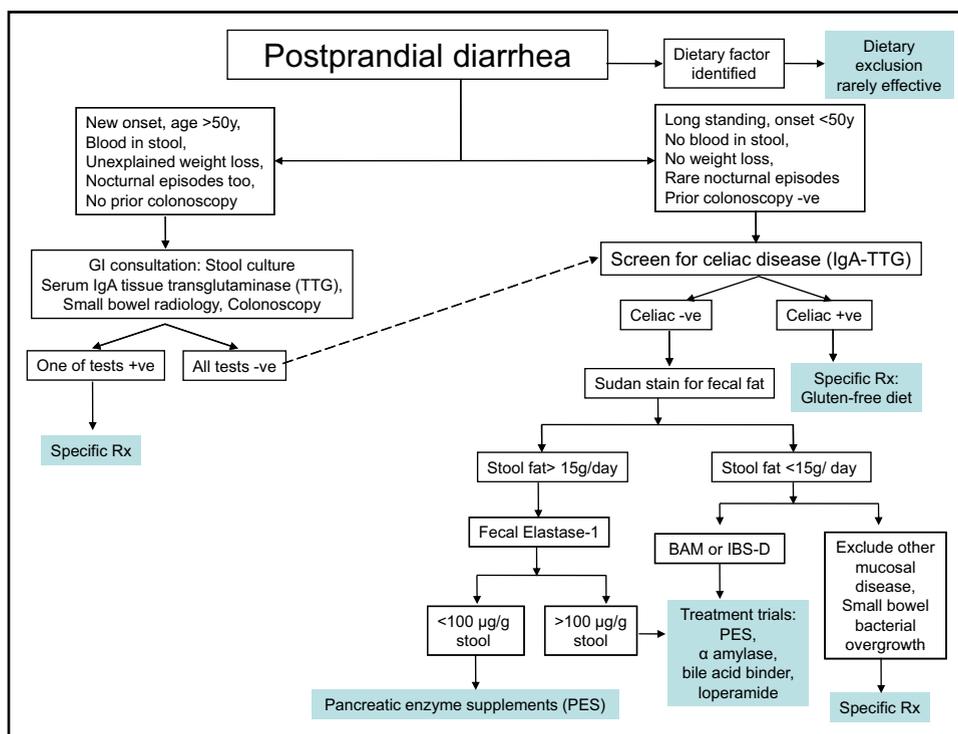


Figure Management and treatment algorithm for postprandial diarrhea. BAM = bile acid malabsorption; GI = gastrointestinal; IBS-D = irritable bowel syndrome-diarrhea; IgA = immunoglobulin A.

CONCLUSION AND RECOMMENDATIONS

Using only symptom-based criteria to establish the diagnosis of irritable bowel syndrome-diarrhea and failing to ask about the relationship of symptoms to eating currently limit physicians. We recommend that primary care physicians focus on the history: when a patient clearly identifies that their episodes of diarrhea occur after eating, a maldigestion condition also should be considered after exclusion of celiac and Crohn disease (Figure). Stool specimens could be analyzed for fecal elastase-1 and fat, not only because these may identify maldigestive process with moderate sensitivity, but because a negative test is useful to exclude the need for further consideration of maldigestive disorders.

Effective treatment with PES or bile acid binders may serve as a surrogate diagnostic test for maldigestion due to bile acid malabsorption, pancreatic or glucosidase insufficiency or inhibition. Current diagnostic testing for any of these conditions is not readily available.

If neither enzyme supplementation nor bile acid sequestrants help, then treatment for small bowel overgrowth is reasonable, followed by testing for rare endocrine tumors, (which account for <1% of cases for chronic diarrhea) such as carcinoid, medullary thyroid cancer, systemic mastocytosis, gastrinomas, VIPomas, glucagonomas, and somatostatinomas.⁶⁹

More research is required to better understand this postprandial diarrhea syndrome. However, there is enough known to conclude that to include these maldigestive conditions

reviewed as simply irritable bowel syndrome-diarrhea is no longer appropriate, as it deprives the patient of a potential specific and effective treatment.

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References

- Manning AP, Thompson WG, Heaton KW, Morris AF. Towards positive diagnosis of the irritable bowel. *Br Med J.* 1978;2 (6138): 653-654.
- Lacy BE, Weiser K, De Lee R. The treatment of irritable bowel syndrome. *Therap Adv Gastroenterol.* 2009;2(4):221-238.
- Jellema P, van der Windt DA, Schellevis FG, van der Horst HE. Systematic review: accuracy of symptom-based criteria for diagnosis of irritable bowel syndrome in primary care. *Aliment Pharmacol Ther.* 2009;30(7):695-706.
- Ragnarsson G, Bodemar G. Pain is temporally related to eating but not to defaecation in the irritable bowel syndrome (IBS). Patients' description of diarrhea, constipation and symptom variation during a prospective 6-week study. *Eur J Gastroenterol Hepatol.* 1998;10(5):415-421.
- Steed KP, Bohemen EK, Lamont GM, Evans DF, Wilson CG, Spiller RC. Proximal colonic response and gastrointestinal transit after high and low fat meals. *Dig Dis Sci.* 1993;38(10):1793-1800.
- Deiteren A, Camilleri M, Bharucha AE, et al. Performance characteristics of scintigraphic colon transit measurement in health and irritable bowel syndrome and relationship to bowel functions. *Neurogastroenterol Motil.* 2010;22(4):415-423, e495.

7. Marciari L, Cox EF, Hoad CL, et al. Postprandial changes in small bowel water content in healthy subjects and patients with irritable bowel syndrome. *Gastroenterology*. 2010;138(2):469-477. 477.e1.
8. Camilleri M. Do the symptom-based, Rome criteria of irritable bowel syndrome lead to better diagnosis and treatment outcomes? The con argument. *Clin Gastroenterol Hepatol*. 2009;8(2):129.
9. Gudmand-Hoyer E, Skovbjerg H. Disaccharide digestion and mal-digestion. *Scand J Gastroenterol Suppl*. 1996;216:111-121.
10. Gonlachanvit S, Mahayosnond A, Kullavanijaya P. Effects of chili on postprandial gastrointestinal symptoms in diarrhoea predominant irritable bowel syndrome: evidence for capsaicin-sensitive visceral nociception hypersensitivity. *Neurogastroenterol Motil*. 2009;21(1): 23-32.
11. Floch MH, Narayan R. Diet in the irritable bowel syndrome. *J Clin Gastroenterol*. 2002;35 (1 Suppl):S45-S52.
12. Whorwell P, Lea R. Dietary treatment of the irritable bowel syndrome. *Curr Treat Options Gastroenterol*. 2004;7(4):307-316.
13. Fernandez-Banares F, Esteve M, Viver JM. Fructose-sorbitol malab-sorption. *Curr Gastroenterol Rep*. 2009;11(5):368-374.
14. Zar S, Benson MJ, Kumar D. Food-specific serum IgG4 and IgE titers to common food antigens in irritable bowel syndrome. *Am J Gastro-entrol*. 2005;100(7):1550-1557.
15. Drisko J, Bischoff B, Hall M, McCallum R. Treating irritable bowel syndrome with a food elimination diet followed by food challenge and probiotics. *J Am Coll Nutr*. 2006;25(6):514-522.
16. McKee AM, Prior A, Whorwell PJ. Exclusion diets in irritable bowel syndrome: are they worthwhile? *J Clin Gastroenterol*. 1987;9(5):526-528.
17. Kanazawa M, Fukudo S. Effects of fasting therapy on irritable bowel syndrome. *Int J Behav Med*. 2006;13(3):214-220.
18. Leeds JS, Hopper AD, Sidhu R, et al. Some patients with irritable bowel syndrome may have exocrine pancreatic insufficiency. *Clin Gastroenterol Hepatol*. 2010;8(5):433-438.
19. Hardt PD, Krauss A, Bretz L, et al. Pancreatic exocrine function in patients with type 1 and type 2 diabetes mellitus. *Acta Diabetol*. 2000;37(3):105-110.
20. Ewald N, Bretzel R, Fantus I, et al. Pancreatin therapy in patients with insulin-treated diabetes mellitus and exocrine pancreatic insufficiency according to low fecal elastase 1 concentrations. Results of a prospec-tive multi-centre trial. *Diabetes Metab Res Rev*. 2007;23(5):386-391.
21. Hahn J, Kerner W, Maisonneuve P, Lowenfels A, Lankisch P. Low fecal elastase 1 levels do not indicate exocrine pancreatic insufficiency in type-1 diabetes mellitus. *Pancreas*. 2008;36(3):274-278.
22. Camilleri M. Integrated upper gastrointestinal response to food intake. *Gastroenterology*. 2006;131(2):640-658.
23. Suarez FL, Savaiano DA, Levitt MD. A comparison of symptoms after the consumption of milk or lactose-hydrolyzed milk by people with self-reported severe lactose intolerance. *N Engl J Med*. 1995;333(1): 1-4.
24. Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. *Oxid Med Cell Longev*. 2009;2(5):270-278.
25. Tundis R, Loizzo MR, Menichini F. Natural products as alpha-amylase and alpha-glucosidase inhibitors and their hypoglycaemic potential in the treatment of diabetes: an update. *Mini Rev Med Chem*. 2010;10(4): 315-331.
26. Hanhineva K, Torronen R, Bondia-Pons I, et al. Impact of dietary polyphenols on carbohydrate metabolism. *Int J Mol Sci*. 2010;11(4): 1365-1402.
27. Shihabudeen MS, Priscilla H, Thirumurugan K. Cinnamon extract inhibits alpha-glucosidase activity and dampens postprandial glucose excursion in diabetic rats. *Nutr Metab (Lond)*. 2011;8(1):46.
28. Oboh G, Ademiluyi AO, Faloye YM. Effect of combination on the antioxidant and inhibitory properties of tropical pepper varieties against α -amylase and α -glucosidase activities in vitro. *J Med Food*. 2011;14(10):1152-1158.
29. El-Beshbishy HA, Bahashwan SA. Hypoglycemic effect of basil (*Oci-mum basilicum*) aqueous extract is mediated through inhibition of α -glucosidase and α -amylase activities: an in vitro study. *Toxicol Ind Health*. 2011 Jun 2 [Epub ahead of print].
30. Lee CH, Garcia HS, Parkin KL. Bioactivities of kernel extracts of 18 strains of maize (*Zea mays*). *J Food Sci*. 2010;75(8):C667-C672.
31. Patil SB, Ghadyale VA, Taklikar SS, Kulkarni CR, Arvindekar AU. Insulin secretagogue, alpha-glucosidase and antioxidant activity of some selected spices in streptozotocin-induced diabetic rats. *Plant Foods Hum Nutr*. 2011;66(1):85-90.
32. Kang MJ, Kim JH, Choi HN, et al. Hypoglycemic effects of Welsh onion in an animal model of diabetes mellitus. *Nutr Res Pract*. 2010; 4(6):486-491.
33. Konno S, Tortorelis DG, Fullerton SA, Samadi AA, Hettiarachchi J, Tazaki H. A possible hypoglycaemic effect of maitake mushroom on Type 2 diabetic patients. *Diabet Med*. 2001;18(12):1010.
34. Kato A, Minoshima Y, Yamamoto J, Adachi I, Watson AA, Nash RJ. Protective effects of dietary chamomile tea on diabetic complications. *J Agric Food Chem*. 2008;56(17):8206-8211.
35. Chiasson JL, Josse RG, Gomis R, Hanefeld M, Karasik A, Laakso M. Acarbose treatment and the risk of cardiovascular disease and hyper-tension in patients with impaired glucose tolerance: the STOP-NIDDM trial. *JAMA*. 2003;290(4):486-494.
36. Li Y, Zhou F, Gao F, Bian J, Shan F. Comparative evaluation of quercetin, isoquercetin and rutin as inhibitors of alpha-glucosidase. *J Agric Food Chem*. 2009;57(24):11463-11468.
37. Robayo-Torres CC, Quezada-Calvillo R, Nichols BL. Disaccharide digestion: clinical and molecular aspects. *Clin Gastroenterol Hepatol*. 2006;4(3):276-287.
38. Sander P, Alfalah M, Keiser M, et al. Novel mutations in the human sucrase-isomaltase gene (SI) that cause congenital carbohydrate mal-absorption. *Hum Mutat*. 2006;27(1):119.
39. Leberthal E, Khin Maung U, Zheng BY, Lu RB, Lerner A. Small intestinal glucoamylase deficiency and starch malabsorption: a newly recognized alpha-glucosidase deficiency in children. *J Pediatr*. 1994; 124(4):541-546.
40. Karnsakul W, Luginbuehl U, Hahn D, et al. Disaccharidase activities in dyspeptic children: biochemical and molecular investigations of maltase-glucoamylase activity. *J Pediatr Gastroenterol Nutr*. 2002; 35(4):551-556.
41. Ringrose RE, Preiser H, Welsh JD. Sucrase-isomaltase (palatinase) deficiency diagnosed during adulthood. *Dig Dis Sci*. 1980;25(5):384-387.
42. Sonntag WM, Brill ML, Troyer WG Jr, Welsh JD, Semenza G, Prader A. Sucrose-isomaltase malabsorption in an adult woman. *Gastroen-terology*. 1964;47:18-25.
43. McNair A, Gudmand-Hoyer E, Jarnum S, Orrild L. Sucrose malab-sorption in Greenland. *Br Med J*. 1972;2(5804):19-21.
44. Simadibrata M, Wanders RJ, Jan G, et al. Examination of small bowel enzymes in chronic diarrhea. *J Gastroenterol Hepatol*. 2003;18(1): 53-56.
45. Lifshitz F, Holman GH. Disaccharidase deficiencies with steatorrhea. *J Pediatr*. 1964;64:34-44.
46. Hammer HF, Santa Ana CA, Schiller LR, Fordtran JS. Studies of osmotic diarrhea induced in normal subjects by ingestion of polyeth-ylene glycol and lactulose. *J Clin Invest*. 1989;84(4):1056-1062.
47. Austin GL, Dalton CB, Hu Y, et al. A very low-carbohydrate diet improves symptoms and quality of life in diarrhea-predominant irrita-ble bowel syndrome. *Clin Gastroenterol Hepatol*. 2009;7:706-708.
48. Walters JR. Defining primary bile acid diarrhea: making the diagnosis and recognizing the disorder. *Expert Rev Gastroenterol Hepatol*. 2010; 4(5):561-567.
49. Kurien M, Evans KE, Leeds JS, Hopper AD, Harris A, Sanders DS. Bile acid malabsorption: an under-investigated differential diagnosis in patients presenting with diarrhea predominant irritable bowel syn-drome type symptoms. *Scand J Gastroenterol*. 2011;46(7-8):818-822.
50. Walters JR, Pattni SS. Managing bile acid diarrhoea. *Therap Adv Gastroenterol*. 2010;3(6):349-357.

51. Takahama U, Hirota S. Inhibition of buckwheat starch digestion by the formation of starch/bile salts complexes: possibility of its occurrence in the intestine. *J Agric Food Chem*. 2011;59:6277-6283.
52. Suarez F, Levitt M, Adshead J, Barkin J. Pancreatic supplements reduce symptomatic response of healthy subjects to a high fat meal. *Dig Dis Sci*. 1999;44(7):1317-1321.
53. Weber A, Roy C, Chartrand L, et al. Relationship between bile acid malabsorption and pancreatic insufficiency in cystic fibrosis. *Gut*. 1976;17(4):295-299.
54. Boyle B, Long W, Balistreri W, Widzer S, Huang N. Effect of cimetidine and pancreatic enzymes on serum and fecal bile acids and fat absorption in cystic fibrosis. *Gastroenterology*. 1980;78(5 Pt 1): 950-953.
55. Dutta S, Anand K, Gadacz T. Bile salt malabsorption in pancreatic insufficiency secondary to alcoholic pancreatitis. *Gastroenterology*. 1986;91(5):1243-1249.
56. Stern RC, Eisenberg JD, Wagener JS, et al. A comparison of the efficacy and tolerance of pancrelipase and placebo in the treatment of steatorrhea in cystic fibrosis patients with clinical exocrine pancreatic insufficiency. *Am J Gastroenterol*. 2000;95(8):1932-1938.
57. Carroccio A GA, Zuin G. Efficacy of oral pancreatic enzyme therapy for the treatment of fat malabsorption in HIV-infected patients. *Aliment Pharmacol Ther*. 2001;15:1619-1625.
58. Money ME, Hofmann AF, Hagey LR, Walkowiak J, Talley NJ. Treatment of irritable bowel syndrome-diarrhea with pancrealipase or colessevelam and association with steatorrhea. *Pancreas*. 2009;38(2):232-233.
59. Money M, Walkowiak J, Virgilio C, Talley N. Pilot study: a randomized, double blind, placebo controlled trial of pancrealipase for the treatment of postprandial irritable bowel syndrome-diarrhea. *Frontline Gastroenterol*. 2011;2(1):48-56.
60. Nichols BL, Quezada-Calvillo R, Robayo-Torres CC, et al. Mucosal maltase-glucoamylase plays a crucial role in starch digestion and prandial glucose homeostasis of mice. *J Nutr*. 2009;139(4):684-690.
61. Hammer HF, Fine KD, Santa Ana CA, Porter JL, Schiller LR, Fordtran JS. Carbohydrate malabsorption. Its measurement and its contribution to diarrhea. *J Clin Invest*. 1990;86(6):1936-1944.
62. Malfertheiner P, Dominguez-Munoz JE. Effect of exogenous pancreatic enzymes on gastrointestinal and pancreatic hormone release and gastrointestinal motility. *Digestion*. 1993;54(Suppl 2):15-20.
63. Layer P, Keller J. Pancreatic enzymes: secretion and luminal nutrient digestion in health and disease. *J Clin Gastroenterol*. 1999;28(1):3-10.
64. Wedlake L, A'Hern R, Russell D, Thomas K, Walters JR, Andreyev HJ. Systematic review: the prevalence of idiopathic bile acid malabsorption as diagnosed by SeHCAT scanning in patients with diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther*. 2009;30(7):707-717.
65. Habba SF. Diarrhea predominant irritable bowel syndrome (IBS-D): fact or fiction. *Med Hypotheses*. 2011;76(1):97-99.
66. Odunsi-Shiyabade ST, Camilleri M, McKinzie S, et al. Effects of chenodeoxycholate and a bile acid sequestrant, colessevelam, on intestinal transit and bowel function. *Clin Gastroenterol Hepatol*. 2010; 8(2):159-165.
67. Spiller GA, Story JA, Lodics TA, et al. Effect of sun-dried raisins on bile acid excretion, intestinal transit time, and fecal weight: a dose-response study. *J Med Food*. 2003;6(2):87-91.
68. Vitali F, Bonina FP, Saija A, et al. Studies on antidiarrhoeal activity of an extract of wine from Jacquez grapes in mice. *Phytother Res*. 2005;19(11):924-927.
69. Jensen RT. Pancreatic endocrine tumors: recent advances. *Ann Oncol*. 1999;10(Suppl 4):170-176.